Topical Reviews

Oncogenes and onco-suppressor genes in lung cancer

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Introduction

It has been known for many years that the evolution of cancer is a multi-step process [for review see (1) and references therein]. Insight into the nature of some of the steps has been gained through advances in molecular and cellular biology and it has become apparent that mutations in a limited number of genes which control cell proliferation and differentiation is a key feature of this process. Two categories of genes in particular have been implicated: proto-oncogenes and tumour-suppressor genes [for review see (2)]. Protooncogenes become 'activated' to oncogenes through particular mutations so that either the normal protein is overexpressed or a mutant protein is produced. Expression of tumour suppressor genes is believed to restrain cells from unregulated proliferation and inactivation of both homologues releases cells from these controls.

Numerous studies have been undertaken to determine the mutational events that are key to particular cancers. For colon cancer, it has been shown that the same type of tumour can arise through activation of different sets of genes (3,4). This is also likely to be true for other cancers including pulmonary carcinoma.

Lung cancer is one of the commonest cancers in the developed world for both men and women. Prognosis is poor with fewer than 13% of all lung cancer patients living more than 5 yr after diagnosis [for reviews see (5,6)]. With such an appalling mortality it is important to elucidate the genetic changes involved in the pathogenesis of lung cancer and to determine how these changes correlate with clinical features. More detailed information may allow earlier diagnosis, better tumour typing, more focused treatment and improved prognosis.

Lung cancers are divided into two main groups on the basis of their histology (5). The first group, small cell lung cancer (SCLC) accounts for about 25% of all lung tumours, is an extremely aggressive neoplasm, is frequently associated with distant metastases and has the poorest prognosis of all lung tumours. It was formerly treated by surgery, but is now treated primarily with chemotherapy and radiotherapy which give slightly improved survival rates (7). The second group, comprising the remaining 75% of lung cancers, are non-small cell lung cancers (NSCLC). There are several sub-types which are characterized by their tissue of origin: adenocarcinomas which arise in the major bronchi, squamous carcinomas which arise in the squamous epithelium and large cell carcinomas which are largely undifferentiated tumours probably arising from stem or basal cells. NSCLC are treated primarily with surgery and radiotherapy. SCLC and NSCLC are regarded as being separate clinicopathological entities on the basis of their morphology, sensitivity to chemotherapeutic agents and ionizing radiation. They will be treated separately in this review.

Oncogenes in Non Small Cell Lung Cancer

RAS

The K-ras, H-ras and N-ras genes encode highly homologous protein monomers, $p21^{ras}$, that are located on the cytoplasmic side of the plasma membrane. Their similarity in sequence to G proteins suggests that they may play a role in transmitting proliferation signals from receptors at the cell surface to the nucleus (8). They bind guanosine triphosphate (GTP) and display an intrinsic GTPase activity that is enhanced by interaction with the protein, GAP (9). Ras proteins are thought to be in an active state for signalling when GTP is bound and inactive when the GTP has been hydrolysed to GDP (8). Overexpression of the normal gene is activating *in vitro* (8) as are point mutations in codons 12, 13 or 61 which prevent intrinsic GTP hydrolysis presumably by trapping the protein in the active form (10).

Amplification of the *ras* genes is uncommon in NSCLC (11–13). One study could find no evidence for high levels of overexpression of *ras* (14), however, another group found elevated levels of $p21^{ras}$ proteins compared to normal adjacent tissue from the same patient (15–17). The level and frequency varies with the tumour type with about 80% of adenocarcinomas compared to 50% of squamous cell carcinomas displaying increased levels of *ras* p21. The exact nature of the protein(s) detected is unknown because the monoclonal antibody used (Y13-259) in these studies neither distinguishes between H-, K- and N-RAS proteins nor whether the protein is normal or mutant.

Activating mutations in H-ras and N-ras genes are rare in NSCLCs (11,18) except perhaps in Japan (19). There is now evidence linking mutations in any member of the ras family with poor prognosis (20). K-ras is mutated in approximately 30% of adenocarcinomas, but this activation is rare in other types of NSCLC (13.21). Several groups have now shown that there is a correlation between K-ras mutations in adenocarcinomas and a history of smoking (21-24) with about 30% of smokers compared with 2% of non smokers having G to T transversions at codon 12 (21-23). This type of mutation is consistent with exposure of the lung to carcinogens in tobacco smoke such as benzo[a] pyrene. No correlation was found between K-ras mutations and sex, age at diagnosis or tumour stage (21), but mutations in K-ras are associated with a very poor prognosis (20,21,25).

MYC

The myc gene family encodes at least three proteins c-MYC, N-MYC and L-MYC, of M_r 62–68 kDa. [For review see (26)]. The protein products are located in the nucleus, bind to DNA and associate with at least one other protein, Max (27). The function of MYC proteins has not been established, but they may be transcriptional factors (28). Activation of myc is thought to occur by gene amplification or overexpression of the protein, but not by point mutations.

Amplification of the *c-myc* gene is found in about 10% and over expression in about 50% of NSCLCs of all types (15). L-myc and N-myc appear not to be activated in NSCLC although it has been reported that N-myc is amplified in a pulmonary adenocarcinoma cell line (29).

Recent RFLP studies have shown that in Japanese patients with lung cancer, the form of the L-myc gene correlates with prognosis, metastasis and the incidence of multiple cancers (30,31). This correlation is particularly strong for adenocarcinoma and squamous-cell

carcinoma. However, such correlations were not found in studies of Norwegian patients (32) or of white and black patients (33). The reason for these contradictory results is not known, but may arise from differences in the type of tumour analysed or in the ethnic origin of patients.

ErbB-1

ErbB-1 encodes the epidermal growth factor receptor, a transmembrane glycoprotein of M_r 170 kDa that possesses intrinsic tyrosine kinase activity and is though to play a role in signal transduction [for review see (34)]. Activation is commonly by overexpression of the protein, but not by mutation (35).

Few studies have been undertaken on the expression of *erbB-1* in lung cancer, but it is reported that about 25% of squamous SCLCs overexpress *erbB-1* protein (36).

C-erbB-2

The *c-erbB-2 (neu)* gene encodes a transmembrane, tyrosine-specific protein kinase, $p185^{neu}$. It is a putative growth factor receptor, related in sequence and structure to the epidermal growth factor receptor (37). The gene is frequently amplified in adenocarcinomas and mRNA and protein levels are frequently elevated in tumours as compared to normal tissue.

Amplification of the *c-erbB-2* proto-oncogene is an infrequent event in lung cancer (38). However, overexpression of p185^{neu} as detected immunohistochemically, is frequently found in NSCLCs – 10/29 adenocarcinomas and 5/16 squamous cell carcinomas (39). In adenocarcinomas, p185^{neu} expression tended to be found in older patients and there was an independent association with short survival times. In squamous cell carcinomas, p185^{neu} expression did not correlate with these parameters.

C-FOS AND C-JUN

c-fos and c-jun encode nuclear proteins that form a complex, AP-1, which acts as a transcriptional factor for genes possessing a specific DNA recognition site (40). Recently, it was reported that in adeno- and squamous cell lung carcinomas, the level of AP-1 activity in nuclear extracts was elevated in ten out of 13 tumours as compared to their adjacent normal tissues (41). As there is evidence that AP-1 may be involved in signal transduction (42), elevated levels may be oncogenic. It has not been established if changes in the AP-1 levels in a tumour correlate with prognosis.

C-RAF-I

The c-raf proto-oncogene encodes a serine/threoninespecific protein kinase, p74^{raf-1}, which is located on the internal side of the plasma membrane (43). RFLP analyses of 73 unmatched NSCLC tumours showed that there was loss of heterozygosity in 31 suggesting that this gene might play a role in the pathogenesis of the cancer (44).

С-МҮВ

The c-myb proto-oncogene encodes a nuclear protein of 75 kDa which is a transcriptional regulator (45). DNA hydridization studies showed that three out of four adenocarcinomas had lost heterozygosity for c-myb (38). Analyses of several NSCLCs have shown defects in RNA transcription. Thus in 11 NSCLC cell lines no c-myb transcripts were detected (46). These results may indicate that aberrant c-myb expression may play a role in generation of lung cancer.

Onco-suppressor Genes in Non Small Cell Lung Cancer

P53

The p53 gene encodes a 53-kDa nuclear phosphoprotein which has recently been identified as a transcriptional activator (47). High levels of the wild type gene product inhibit growth possibly through acting as a checkpoint for DNA damage at the G0–G1 transition of cell division (48,49). Several mutant alleles with particular single base substitutions encode proteins that confer altered cell growth regulatory properties (48).

Mutations in p53 are the commonest genetic changes detected in several different types of cancers and are a common feature of NSCLCs (50). The frequency varies with the type of pulmonary cancer with about 67% of squamous cell carcinomas and 37% of adenocarcinomas carrying p53 mutations (51,52). Mutations have also been reported in large cell lung cancers, three out of six, but the number of cases studied was low (52,53). No statistically significant correlation has been found between p53 mutations and the age or sex of patients or the histology, clinical stage, or lymph node involvement (51).

G:C to T:A transversions are found in about 50% of NSCLCs (51,54). This is a type of mutation in p53 that is uncommon in other types of human cancer (50). Since one of the components of cigarette smoke is benzo[a] pyrene, a potent mutagen that causes G:C to T:A transversions (55), a correlation between smoking and these particular mutations might be expected. While a study of NSCLCs in Japanese patients showed that there is indeed a statistically significant association of p53 mutations with lifetime consumption of cigarettes (51), an earlier analysis of p53 mutations in American lung cancer patients found a different nucleotide substitution pattern and failed to find any significant correlation between smoking and p53 mutations (52). The reasons for these contradictory results are unknown. One possibility is that there may be differences in the genetic susceptibility to lung cancer in the two populations.

Mutations at many positions along the p53 gene have been found (51,53,56). Codon 273 has been reported to be a 'hot spot' for mutation in American patients with lung cancer, but none of 26 patients carried a mutation at this site in a Japanese study (51).

Immunocytochemical studies have shown higher levels of mutant p53 protein in squamous cell and in adenocarcinomas of the lung than in normal adjacent tissue (17). The higher the level of p53 present, the poorer was the prognosis both with respect to the primary tumour (P < 0.05) and to lymph node metastases (P < 0.005) (17).

p53 alterations have been detected in preneoplastic lesions of the lung (57,58) suggesting that p53 changes occur in the early stages of lung cancer development. The possibility arises that screening for p53 mutations could form the basis for early detection of lung cancer.

p53 binds to the product of the MDM2 gene (59) which was originally identified as a murine dominant transforming oncogene (60). The MDM2 gene maps to 12q13–14 in man, a chromosomal region that is often altered in sarcomas (61). Amplification of the MDM2 gene is found in over one third of sarcomas (59) and it has been suggested that in cancers where only nonmutated p53 can be detected, functional p53 protein may nevertheless be unavailable to the cell by virtue of being sequestered by excess MDM2 (59). The status of the MDM2 gene in lung cancer will doubtless be the subject of intense investigation.

Rв

The retinoblastoma (Rb-1) gene encodes a DNA binding protein of 110 kDa that is thought to be involved in events important in cell division (48). Inactivation of the Rb gene by deletion and loss of heterozygosity have been found in several cancers (48). The status of the Rb-1 gene in NSCLC is not known, but no alterations were found in 20 NSCLC-derived cell lines (62). Anti RB1 peptide antibodies precipitated RB1 protein in eight out of nine NSCLC-derived cell lines, so at least in cell lines RB1 protein is commonly expressed (62).

NM23

The nm23 gene located on the long arm of chromosome 17 at 17p11-q11 (63) encodes a nucleoside diphosphate kinase (65). Low levels of nm23 mRNA and the corresponding protein have been found to correlate with high metastatic potential in several tumours (66,67). Somatic deletion of an allele of nm23 has been reported in adenocarcinomas of the lung (63), but an investigation into its pronostic value showed there was no correlation between nm23 product and survival rates (68).

Chromosomal Abnormalities

The presence of consistent chromosomal losses or deletions has suggested the involvement of known or candidate tumour suppressor genes at these locations. This has been confirmed by RFLP analysis showing LOH for genes on chromosomes 13q (RB) and 17p (p53) (69).

The karyotype of NSCLC is complex even in cells obtained from tumours prior to chemotherapy (70). In one recent study, chromosomes 1, 3, 6, 7, 8, 11, 13, 15, 17 and 19 were found to carry non random rearrangements (70). The most frequently rearranged bands being 1p13, 3p13, 8p11–q11, 15p11-q11 and 17p11 each of which appeared in 8-14/30 samples. In a different study RFLP deletion analyses of DNA from 53 primary NSCLCs showed loss of heterozygosity (LOH) to be a frequent event on the long arms of chromosomes 1 (37%), 2 (31%), 5 (30%), 8 (31%), and 13 (32%) and the short arms of chromosomes 3 (54%) and 17 (62%) (71). LOH on chromosomes 3p and 17q were more frequent in squamous- than in adenocarcinomas.

Deletions in the short arm of chromosome 3 occur in over 75% of NSCLCs (72–74). The regions, 3p21.3 and 3p14.1–21.1 were preferentially deleted in adenocarcinomas and loss of heterozygosity at these regions was seen more frequently in poorly- or undifferentiatedthan in well-differentiated tumours (74). In addition, the frequency of deletion was higher in stage III than in stage I or II tumours (P = 0.043). No correlation was found with parameters such as smoking status, node size and distant metastases. In all squamous cell lung carcinomas analysed, there was a deletion at one or both of the regions. These results suggest oncosuppressor genes may be located at 3p21.3 and 3p14.1– 21.1 and that they may be involved in the progression of the tumours.

Changes in Small Cell Lung Carcinoma

Mutations in *ras* genes are essentially absent in SCLC – 0/42 (75), 0/12 (21) and elevation of *ras* protein expression is rare and not of statistical significance (P = 0.07) (17).

Initial studies on *myc* expression in SCLC were performed on cell lines and found evidence for *c-myc*

or N-myc gene amplification (16,17) in about 30% of cell lines. c-myc amplification was seen only in patients who had undergone chemotherapy and a correlation between c-myc amplification in treated patients and shorter survival times was found (17).

Gene amplification of all three members of the mycfamily have been observed in primary SCLC (78,79). In one study 19/26 patients with metastatic SCLC had amplification of either c-myc or N-myc and there was no association between mutations and variant morphology of tumours. Metastases and tumours had similar copy numbers of the genes suggesting that myc amplification was an early event. There does not appear to be a correlation between the degree of mycgene amplification in primary tumour and survival time (79). However, it is necessary to interpret these results with caution as the survival time of SCLC patients after diagnosis is so short anyway.

In accord with data from cell lines, amplification of myc DNA occurs more frequently in patients who have undergone chemotherapy, but cell lines established from SCLCs before and after chemotherapy did not alter their status of myc gene copy number (80). Thus, it is not clear if chemotherapy can actually cause myc gene amplification.

Elevated expression of myc has been observed in five of six patient tumours, but the level of expression did not necessarily correlate with the degree of myc gene amplification (81). In another study of 15 primary biopsies from patients who had not undergone chemotherapy, N-myc RNA levels were increased in six cases. Increased expression of N-myc correlated with a poor subsequent response to chemotherapy, rapid tumour growth and short survival times (P < 0.01) (82). The basis for these observations is not known.

In a study which included 21 patients with SCLC, immunohistochemical analyses showed that c-MYC protein was over-expressed in 4 patients. While the sample number is low this may be of statistical significance (P < 0.05) (83). The status of c-myc amplification did not correlate with prognosis of the patients.

RFLP analyses of 84 primary human lung carcinomas indicated LOH for the c-*raf*-1 locus in five of five informative matched normal and tumour SCLC samples and in 42 of 42 unmatched SCLC tumours (44).

Onco-suppressor Genes in SCLC

Mutations in p53 are present in over 75% of SCLCs (54,84–86). In one study, loss of one p53 allele and mutation of the other was found in 16/16 stage III–IV tumours and 3/6 stage I–II tumours (85). In addition, the allelic loss and/or mutation found in primary

tumours was also observed in metastases in distant organs suggesting that alterations to p53 are early events in the pathogenesis of SCLC.

Structural abnormalities within the Rb gene have been detected in one of eight (13%) primary SCLC tumours and 4/22 (18%) SCLC lines (87,88). Absence of RB mRNA and p105 RB protein however, is common in SCLC lines (87,89).

Deletions of the short arm of chromosome 3 are common events in SCLC (90,91) with three distinct regions between 3p21-25 being involved (73). Four genes, ACY1, APEH, PTPG and D8 have been cloned from this region and their expression in SCLC cell lines compared with levels in normal lung (92-95). All are under-expressed in at least one cell line, but data are so far insufficient to establish a role for any as a tumour suppressor gene in SCLC.

LOH in the tumour suppressor genes, MCC (mutated in colon cancer) and APC (adenomatous polyposis coli), have been implicated in the pathogenesis of several cancers (96–98). Both genes are located on the long arm of chromosome 5 (5q21) a region that is often deleted in SCLC (99). Over 80% of SCLCs show allelic deletion of these genes raising the possibility that they are involved in tumourigenesis (100).

Subtractive hybridization cloning of sequences in a lung cancer and normal cell line led to isolation of three sequences present in normal, but not in the tumour-derived DNA (101). Since one sequence, del-118, was also deleted in a freshly isolated lymph node metastasis from an adenocarcinoma, it is a candidate tumour suppressor gene for lung tumours.

Concluding Remarks

Regardless of the tissue or origin, the development of a cancer cell from a normal cell involves a series of genetic changes that contribute to a loss of normal growth control mechanisms. For lung cancer we are still far from understanding what these events are. Significant progress has been made recently in determining the status of oncogenes such as ras and myc and onco-suppressor genes such as p53 whose role is considered to be important in the multistep process of carcinogenesis. Consistent chromosome deletions may facilitate identification of more tumour suppressor genes and it is worth noting that microscopically visible deletions cover several million base pairs so several tumour suppressor genes could be located 'close' together. The expression in lung cancer patients of neural cell adhesion molecule and blood group antigen A has been linked to favourable prognosis of patients (102,103). This presents another promising line of investigation.

Further detailed studies are required to probe relationships between mutations and factors such as diagnosis: clinical parameters: genetic predisposition and prognosis.

In addition, it will be necessary to try to order the specific genetic alterations in terms of time as this may allow the development of new diagnostic tools, and treatments for lung cancer.

References

- Stein WD. Analysis of cancer incidence data on the basis of multistage and clonal growth models. Adv Cancer Res 1991; 56: 161-213.
- Spandidos DA, Anderson MLM. Oncogenes and oncosuppressor genes. Their involvement in cancer. J Pathol 1989; 157: 1-10.
- Vogelstein B, Fearon ER, Hamilton SR et al. Genetic alterations during colorectal-tumour development. N Engl J Med 1988; 319: 525-532.
- Fearon F, Vogelstein B. A genetic model for colorectal tumorogenesis Cell 1990; 61: 759-767.
- Minna JD, Higgins GA, Glatstern FJ. Cancers of the lung. In: De Vits VT Jr, Hellman S, Rosenberg SA, eds, *Cancer: Principles and Practice of Oncology*. Second edn. Vol. 1. Philadelphia: JB Lipponcott, 1985; 507–597.
- Cancer facts and figures. American Cancer Society: Clifton Road, NE, Atlanta, GA 30329, U.S.A., 1989.
- Bunn PA, Lichter BE, Makuch RW et al. Chemotherapy alone or chemotherapy with chest radiation therapy in limited stage small cell lung cancer: A prospective randomised trial. Ann Intern Med 1987; 106: 655-662.
- Barbacid M. ras genes. Ann Rev Biochem 1987; 56: 799–827.
- Trahey M, McCormick F. A cytoplasmic protein stimulates normal N-ras p21 GTPase, but does not affect oncogenic mutants. Science 1987; 238: 542-545.
- Barbacid M. ras oncogenes: their role in neoplasia. Eur J Clin Invest 1990; 20: 225-235.
- Rodenhuis S, Slebos R, Boot A et al. Incidence and possible clinical significance of K-ras oncogene activation in adenocarcinoma of the human lung. Cancer Res 1988; 48: 5738-5741.
- Slebos RJC, Evers SG, Wagenaar SS, Rodenhuis S. Cellular protooncogenes are infrequently amplified in untreated non-small cell lung cancer (NSCLC). Br J Cancer 1988; 59: 76-80.
- 13. Bos JL. Ras oncogenes in human cancer. Cancer Res 1989; 49: 4682-4689.
- Slebos RJC, Haberts GGM, Evers SG, Mooi WJ, Rodenhuis S. Allele specific detection of K-ras oncogene expression in human non-small cell lung carcinomas. *Int* J Cancer 1991; 48: 51–56.
- Spandidos DA, Zakinthos S, Petraki C et al. Expression of ras p21 and myc p62 oncoproteins in small cell and non small cell carcinoma of the lung. Anticancer Res 1990; 10: 1105–1114.
- Koutselini H, Kappatou G, Yiagnisis M, Field JK, Spandidos DA. Immunocytochemical study of RAS oncoprotein in cytologic specimens of primary lung tumours. *Anticancer Res* 1990; 10: 597-604.
- Papadakis E, Malliri A, Linardopoulos S, Karaiossifidi H, Field JK, Spandidos DA. Ras and p53 expression in

non-small cell lung cancer patients: p53 over-expression correlates with a poor prognosis. *Int J Oncol* 1992; 1: 403-413.

- Reynolds S, Anna CK, Brown KC et al. Activated oncogenes in human lung tumors from smokers. Proc Natl Acad Sci U.S.A. 1991; 88: 1085–1089.
- Suzuki Y, Orita M, Shiraishi M, Hayashi K, Sekiya T. Detection of *ras* gene mutations in human lung cancers by single strand conformation polymorphism analysis of polymerase chain reaction products. *Oncogene* 1990; 5: 1037–1043.
- Mitsudomi T, Steinberg SM, Oie HK et al. ras gene mutations in non-small cell lung cancers are associated with shortened survival irrespective of treatment intent. Cancer Res 1991; 51: 4999-5002.
- Rodenhuis S, Slebos RJC. Clinical significance of ras oncogene activation in human lung cancer. Anticancer Res 1992; 52: 2665–2669.
- Rodenhuis S, van de Wetering M, Mooi W et al. Mutational activation of the K-ras oncogene: a possible pathogenetic factor in adenocarcinoma of the lung. N Engl J Med 1987; 317: 929-935.
- Kobayashi T, Tsuda H, Nogushi M et al. Association of point mutation in c-Ki-ras oncogene in lung adenocarcinoma with particular reference to cytologic subtypes. Cancer 1990; 66: 289–294.
- Slebos RJ, Hruban RH, Dalesio O, Mooi WJ, Offerhaus GJ, Rodenhuis S. Relationship between K-ras oncogene activation and smoking in adenocarcinoma of the human lung. J Natl Cancer Inst 1991; 83: 1024–1027.
- Slebos RJC, Kibbelaar RE, Dalesio O et al. K-ras oncogene activation as a prognostic marker in adenocarcinoma of the lung. N Engl J Med 1990; 323: 561-565.
- Cole MD. The myc oncogene: its role in transformation and differentiation. Ann Rev Genet 1986; 20: 361-384.
- 27. Cole MD. Myc meets its Max. Cell 1991; 65: 715-716.
- Reddy CD, Dasgupta P, Saikumar P, Dudek H, Rauscher III FJ, Reddy EP. Mutational analysis of Max: role of basic, helix-loop-helix/leucine zipper domains in DNA binding, dimerization and regulation of myc-mediated transcriptional activation. Oncogene 1992; 7: 2085-2092.
- Kashii T, Mizushima Y, Nakagawa K, Monno S, Yano S. Amplification of the N-myc oncogene in an adenocarcinoma cell line of the lung. Anticancer Res 1992; 12: 621-624.
- Kawashima K, Shikama H, Imoto K et al. Close correlation between restriction-fragment length polymorphism of the L-myc gene and metastases of human lung cancer to the lymph nodes and other organs. Proc Natl Acad Sci U.S.A. 1988; 85: 2353-2356.
- Kawashima K, Nomura S, Hirai H et al. Correlation of L-myc RFLP with metastasis, prognosis and multiple cancer in lung-cancer patients. Int J Cancer 1992; 50: 557-561.
- Tefre T, Børresen A-L, Aamdal S et al. Studies of the Lmyc DNA polymorphism and relation to metastasis in Norwegian lung cancer patients. B J Cancer 1990; 61: 809-812.
- Tamai S, Sugimira H, Caparoso NE et al. Restrictionfragment-length polymorphism of the L-myc gene locus in a case-control study of lung cancer. Int J Cancer 1990; 46: 411-415.
- 34. Hunter T. The epidermal growth factor receptor gene and its product. *Nature* 1984; 311: 414-424.

- Hendler FJ, Ozanne BW. Human squamous cell line cancers express increased epidermal growth factor receptors. J Clin Invest 1984; 74: 647-651.
- Gorgoulis V, Aninos D, Mikou P et al. Expression of EGF, TGF-a and EGFR in squamous cell lung carcinomas. Anticancer Res 1992; 12: 1183-1188.
- Schecter AL, Stern DF, Vaidyanathan L et al. The NEU oncogene: an erbB related gene encoding a 185 000 M_r tumour antigen. Nature 1984; 312: 513-516.
- Cline MJ, Battifora H. Abormalities of proto-oncogenes in non-small cell lung cancer. Correlations with tumour type and clinical characteristics. *Cancer* 1987; 60: 2669-2674.
- Kern JA, Schwartz DA, Nordberg JE et al. p185^{neu} expression in human lung adenocarcinomas predicts shortened survival. *Cancer Res* 1990; **50**: 5184–5191.
- Ransome LJ, Verma IM. Nuclear proto-oncogenes FOS and JUN. Ann Rev Cell Biol 1990; 60: 539–557.
- 41. Linardopoulos S, Papadakis E, Delakas D, Cranidis A, Spandidos DA. Human lung and bladder carcinoma tumours as compared to their adjacent normal tissue have elevated AP-1 activity at the retinoblastoma gene promoter. *Anticancer Res* 1993; 13: 257–262.
- 42. Vogt PK, Bos TJ. The oncogene jun and nuclear signalling. TIBS 1989; 14: 172-175.
- 43. Morrison DK, Kaplan DR, Rapp U, Roberts TM. Signal transduction from membrane to cytoplasm: growth factors and membrane-bound oncogene products increase raf-1 phosphorylation and associated protein kinase activity. Proc Natl Acad Sci U.S.A. 1988; 85: 8855-8859.
- 44. Sithanandam G, Dean M, Brennscheidt U et al. Loss of heterozygosity at the c-raf locus in small cell lung carcinoma. Oncogene 1989; 4: 451-455.
- 45. Sakura H, Kanei-Ishii C, Nagase T, Nakagoshi H, Gonda TJ, Ishii S. Delineation of three functional domains of the transcriptional activator encoded by the c-myb proto-oncogene. Proc Natl Acad Sci U.S.A. 1989; 86: 5758-5762.
- 46. Kiefer PE, Wegmann B, Bacher M et al. Different pattern of expression of cellular oncogenes in human non-small cell lung cancer cell lines. J Cancer Res Clin Oncol 1990; 116: 29–37.
- Farmer G, Bargonetti J, Zhu H et al. Wild-type p53 activates transcription in vitro. Nature 1992; 358: 83-86.
- Levine AJ, Momand J. Tumor suppressor genes: the p53 and retinoblastoma sensitivity genes and gene products. *Biochem Biophys Acta* 1990; 1032: 119–136.
- Lane DP. p53, guardian of the genome. Nature 1992; 358: 15-16.
- Hollstein M, Sidransky D, Vogelstein B, Harris CC. p53 mutations in human cancers. Science 1991; 253: 49-53.
- Suzuki H, Takahashi T, Kuroishi T et al. p53 mutations in non-small cell lung cancer in Japan: Association between mutations and smoking. Cancer Res 1992; 52: 734-736.
- Chiba I, Takahashi T, Nau MM et al. Mutations in the p53 gene are frequent in primary resected, non-small cell lung cancer. Oncogene 1990; 5: 1603–1610.
- Miller CW, Simon K, Aslo A et al. p53 mutations in human lung tumours. Cancer Res 1992; 52: 1695–1698.
- Takahashi T, Nau MM, Chiba I et al. p53: a frequent target for genetic abnormalities in lung cancer. Science 1989; 246: 491–494.
- Mazur M, Glickman B. Sequence specificity of mutations induced by benzo[a]pyrene-7, 8-diol, 9,10-epoxide at

endogeneous aprt gene in CHO cells. Cell Mol Genet 1988; 14: 393-400.

- Medcalf EA, Takahashi T, Chiba I, Minna J, Milner JH. Temperature-sensitive mutants of *p53* associated with human carcinoma of the lung. *Oncogene* 1992; 7: 71-76.
- Sozzi G, Miozza M, Donghi R et al. Deletions of 17p and p53 mutations in preneoplastic lesions of the lung. *Cancer Res* 1992; 52; 6079-6082.
- Sundaresan V, Ganly P, Hasleton P et al. p53 and chromosome 3 abnormalities, characteristic of malignant lung tumours are detectable in preinvasive lesions of the bronchus. Oncogene 1992; 7: 1989–1997.
- Oliner JD, Kinzler KW, Meltzer PA et al. Amplification of a gene encoding a p53-associated protein in human sarcomas. Nature 1992; 358: 80-83.
- Fakharzadeh SS, Trusko SP, George DL. Tumorigenic potential associated with enhanced expression of a gene that is amplified in a mouse tumor cell line. *EMBO J* 1991; 10: 1565–1569.
- Turc-Carel C, Limon J, Dallin P, Rao U, Karakousis C, Sandberg AA. Cytogenetic studies of adipose tissue tumors II. Recurrent reciprocal translocation t[12:16] [q13:p11] in myxoid liposarcomas. *Cancer Genet Cytogenet* 1991; 23: 291-299.
- Yokota J, Akiyama T, Fung Y-KT et al. Altered expression of the retinoblastoma (Rb) gene in small cell carcinoma of the lung. Oncogene 1988; 3: 471-475.
- Leone A, McBride O, Weston A et al. Somatic deletion of nm23 in human cancer. Cancer Res 1991; 51: 2490-2493.
- Kimura N, Shimada N. Membrane associated nucleotide diphosphate kinase from rat liver. J Biol Chem 1988; 263: 4647-4653.
- 65. Wallett V, Mutzel R, Troll H et al. Dictostelium nucleotide diphosphate kinase highly homologous to nm23 and awd proteins involved in mammalian tumour metastasis and Drosophila development. J Natl Cancer Inst 1990; 82: 1199-1202.
- 66. Steeg PS, Bevilacqua G, Kopper L et al. Evidence for a novel gene associated with low tumour metastatic potential. J Natl Cancer Inst 1988; 80: 200-204.
- 67. Hirayama R, Sawai S, Takagagi Y et al. Positive relationship between expression of anti-metastatic factor (nm23 gene product or nucleotide diphosphate kinase) and good prognosis in human breast cancer. J Natl Cancer Inst 1990; 83: 1249-1250.
- Higashiyama M, Doi O, Yokouchi H et al. Immunohistochemical analaysis of nm23 gene product/NDP kinase expression in pulmonary adenocarcinoma: lack of prognostic value. Br J Cancer 1992; 66: 533-536.
- Takahashi T, Nau MM, Chiba I. p53: a frequent target for genetic abnormality in lung cancer. *Science* 1989; 246: 491-494.
- Testa JR, Siegfried JM. Chromosome abnormalities in human non-small cell lung cancer. *Cancer Res* 1992; 52: 2702s-2706s.
- Tsuchiya E, Nakamara Y, Weng S-Y et al. Allelotype of non-small cell lung carcinoma – comparison between loss of heterozygosity in squamous cell carcinoma and adenocarcinoma. *Cancer Res* 1992; 52: 2478–2481.
- Kok K, Osinga J, Carritt B et al. Deletion of a DNA sequence at the chromosomal region 3p21 in all types of lung cancer. Nature 1987; 330: 578-581.

- Hibi K, Takahashi T, Yamakawa K et al. Three distinct regions involved in 3p deletions in human lung cancer. Oncogene 1992; 7: 445-449.
- 74. Yokoyama S, Yamakawa K, Tsuchiya E et al. Deletion mapping on the short arm of chromosome 3 in squamous cell carcinoma and adenocarcinoma of the lung. *Cancer Res* 1992; 52: 873–877.
- Mitsudomi T, Villet J, Mulshine JL, Linorilla RI, Minna JD, Gazder AF. Mutations of ras genes distinguish a subset of non small cell lung cancer cell lines from small-cell lung cancer lines. Oncogene 1991; 6: 1353-1362.
- Nau MM, Brooks BJ, Carney DN et al. Human small cell lung cancers with amplification and expression of the N-myc gene. Proc Natl Acad Sci U.S.A. 1986; 83: 1092-1096.
- 77. Johnson BE, Ihde DC, Makuch RW et al. Myc family oncogene amplification in tumour cell lines established from small cell lung cancer patients and its relationship to clinical status and course. J. Clin Invest 1987; 79: 1629–1634.
- Wong AJ, Ruppert JM, Eggleton J, Hamilton SR, Baylin SB, Vogelstein B. Gene amplification of c-myc and N-myc in small cell carcinoma of the lung. Science 1986; 23: 461-464.
- Johnson BE, Makuch RW, Simmons AD, Gazder AF, Burch D, Cashell AW. myc family DNA amplification in small cell lung cancer patients' tumors and corresponding cell lines. *Cancer Res* 1988; 48: 5163-5166.
- Brennan J, O'Connor T, Makuch RW et al. myc family DNA amplification in 107 tumors and tumor cell lines from patients with small cell lung cancer treated with different combination chemotherapy regimes. Cancer Res 1991; 51: 1708–1712.
- Takahashi T, Okota Y, Sekido Y et al. Expression and amplification of myc gene family in small cell lung cancer and its relation to biological systems. Cancer Res 1988; 49: 2683-2688.
- Funna K, Steinholz L, Nau E, Bergh J. Increased expression of N-myc in human small cell lung cancer biopsies predicts lack of response to chemotherapy and poor prognosis. Am J Clin Pathol 1987; 88: 216-220.
- Gosney JR, Field JK, Gosney MA, Lye MDW, Spandidos DA, Butt SA. c-myc oncoprotein in bronchial carcinoma: expression in all major morphological types. Anticancer Res 1990; 10: 623-628.
- 84. Iggo R, Gatter K, Bartek J, Lane D, Harris AL. Increased expression of mutant forms of p53 oncogene in primary lung cancer. *Lancet* 1990; **335**: 675–679.
- 85. Sameshima Y, Matsuno Y, Hirohashi S et al. Alterations of the p53 gene are common and critical events for the maintenance of malignant phenotypes in smallcell lung carcinoma. Oncogene 1992; 7: 451-457.
- D'Amico D, Carbone D, Mitsudomi T et al. High frequency of somatically acquired p53 mutations in smallcell lung cancer cell lines and tumors. Oncogene 1992; 7: 339-346.
- Harbour JW, Lai S-L, Whang-Peng J et al. Abnormalities in structure and expression of the human retinoblastoma gene in SCLC. Science 1988; 241: 353–357.
- Horowitz JM, Park S-H, Bogenmann E et al. Frequent inactivation of the retinoblastoma anti-oncogene is restricted to a subset of human tumor cells. Proc Natl Acad Sci U.S.A. 1990; 87: 2775-2779.

- Yokota J, Akujama T, Fung Y-Kt et al. Altered expression of the retinoblastoma (RB) gene in small-cell carcinoma of the lung. Oncogene 1988; 3: 471-475.
- Naylor SL, Johnson BE, Minna JD, Sakaguchi AY. Loss of heterozygosity of chromosome 3p markers in small-cell lung cancer. *Nature* 1987; 329; 451-454.
- Brauch H, Johnson B, Hovis J et al. Molecular analysis of the short arm of chromosome 3 in small-cell and nonsmall-cell carcinoms of the lung. N Engl J Med 1987; 317: 1109–1113.
- Miura I, Graziano SL, Cheng GJ et al. Chromosome alterations in human small cell lung cancer: frequent involvement of 5q. Cancer Res 1992; 52: 1322–1328.
- Carritt B, Kok K, van den Berg A et al. A gene from human chromosome region 3p21 with reduced expression in small cell lung cancer. *Cancer Res* 1992; 52: 1536–1541.
- 94. Naylor SL, Marshall A, Henschel C *et al.* The DNF15S2 locus is transcribed in normal lung and small cell lung cancer. *Genomics* 1989; **4**: 355–361.
- Miller YE, Kao B, Gratzer AF. Lack of expression of aminoacylase-1 in small cell lung cancer. Am J Hum Genet 1987; 46: A32.
- 96. LaForgia S, Morse B, Levy J et al. Receptor-proteintyrosine phosphatase γ is a candidate tumor suppressor gene at human chromosomal region 3p21. Proc Natl Acad Sci U.S.A. 1991; 88: 5036-5040.

- Kintzler KW, Nilbert MC, Su L et al. Identification of a gene located at chromosome 5q21 that is mutated in colon cancers. Science 1991; 253: 661–664.
- Groden J, Thliveris A, Samowitz W et al. Identification and characterisation of the familial adenomatous polyposis coli gene. Cell 1991; 66: 589–600.
- 99. Nishiso I, Nakamura Y, Mitoshi Y et al. Mutations of chromosome 5q21 genes in FAP patients and colorectal cancer patients. Science 1991; 253: 661-664.
- 100. D'Amico D, Carbone DP, Johnson BE et al. Polymorphic sites within the MCC and APC loci reveal very frequent loss of heterozygosity in human small cell lung cancer. Cancer Res 1992; 52: 1996–1999.
- Wieland I, Bohm M, Bogatze S. Isolation of DNA sequences deleted in lung carcinoma by genomic difference cloning. *Proc Natl Acad Sci U.S.A.* 1992; 89: 9705–9709.
- 102. Kibbelaar RE, Moolenaar KEC, Michalides RJAM et al. Neural Cell adhesion molecule expression, neuroendocrine differentiation and prognosis in lung carcinoma. Eur J Cancer 1991; 27: 431–435.
- 103. Lee JS, Ro JY, Sahin AA et al. Expression of bloodgroup antigen A-a favourable prognostic factor in non-small cell lung cancer. New Engl J Med 1991; 324: 1084-1090.